

Package: gedi2 (via r-universe)

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Description A memory-efficient implementation for integrating gene expression data from single-cell RNA sequencing experiments. Uses a C++ backend with thin R wrappers to enable analysis of large-scale single-cell datasets. The package supports multiple data modalities including count matrices, paired data (splicing, RNA velocity, CITE-seq), and binary indicators. It implements a latent variable model with block coordinate descent optimization for dimensionality reduction and batch effect correction. Core algorithms are described in Madrigal et al. (2024) <[doi:10.1038/s41467-024-50963-0](https://doi.org/10.1038/s41467-024-50963-0)>.

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BugReports <https://github.com/csglab/gedi2/issues>

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check_optional_dependencies
Check GEDI Optional Dependencies

Description

Check which optional dependencies are installed and display their status.

Usage

```
check_optional_dependencies()
```

Value

Named logical vector indicating which optional packages are installed

Examples

```
check_optional_dependencies()
```

CreateGEDIOObject *Create GEDI Object*

Description

Creates and configures a GEDI (Gene Expression Data Integration) model object. This implementation uses Option 2 memory optimization: C++ computes Y_i from M , eliminating duplicate Y_i storage in R.

Usage

```
CreateGEDIOObject(
  Samples,
  M = NULL,
  Y = NULL,
  X = NULL,
  colData = NULL,
  C = NULL,
  H = NULL,
  K = 40,
  mode = "Bsphere",
  adjustD = TRUE,
  orthoZ = TRUE,
  Z_shrinkage = 1,
  A_shrinkage = 1,
  Qi_shrinkage = 1,
  Rk_shrinkage = 1,
  oi_shrinkage = 1,
  o_shrinkage = 1,
  si_shrinkage = 1,
  fixed_si = NA,
  rsvd_p = 10,
  rsvd_sdist = "normal",
  verbose = 1,
  num_threads = 1
)
```

Arguments

Samples	Factor or character vector indicating sample of origin for each cell
M	Raw count matrix (sparse or dense), or list of two matrices for paired data. C++ will compute $Y_i = \log(M+1)$ internally - no Y_i copy stored in R!
Y	Log-transformed expression matrix (optional if M provided)
X	Binary indicator matrix (optional if M or Y provided)
colData	Optional data.frame with cell metadata

C	Gene-level prior matrix (genes x pathways)
H	Sample-level covariate matrix (covariates x samples)
K	Number of latent factors (default: 10)
mode	Normalization mode: "B12" or "Bsphere" (default: "B12")
adjustD	Whether to adjust D based on B row norms (default: TRUE)
orthoZ	Whether Z columns should be orthogonal (default: TRUE)
Z_shrinkage, A_shrinkage, Qi_shrinkage, Rk_shrinkage, oi_shrinkage, o_shrinkage, si_shrinkage	Regularization strengths (default: 1)
fixed_si	Fix cell library sizes at this value, or NA to optimize (default: NA)
rsvd_p	Oversampling parameter for randomized SVD (default: 10)
rsvd_sdist	Random distribution for rSVD: "normal", "unif", or "rademacher" (default: "normal")
verbose	Verbosity level: 0 (silent), 1 (info), 2 (debug) (default: 1)
num_threads	Number of OpenMP threads (default: 0 = auto)

Value

GEDI R6 object with memory-efficient architecture

Examples

```
# Load example data
pbmc_small <- SeuratObject::pbmc_small

# Basic usage - memory efficient!
model <- CreateGEDIOobject(
  Samples = pbmc_small@meta.data$orig.ident,
  M = pbmc_small@assays$RNA@counts, # Only M stored in R; Yi computed in C++
  K = 10,
  num_threads = 1
)

# Train the model
model$train(iterations = 50)

# Access results via active bindings
Z <- model$Z
params <- model$params

# Access projections (computed on demand, cached)
zdb <- model$projections$ZDB
db <- model$projections$DB

# Access imputed expression
Y_imputed <- model$imputed$Y() # No M needed!
Y_var <- model$imputed$variance(pbmc_small@assays$RNA@counts) # M required
```

gedi_to_seurat *Convert GEDI Model to Seurat Object*

Description

Creates a Seurat object from a trained GEDI model, including imputed data, projections, and embeddings.

Usage

```
gedi_to_seurat(
  model,
  M = NULL,
  project = "GEDI",
  assay = "RNA",
  use_imputed = TRUE,
  add_projections = TRUE,
  add_embeddings = TRUE,
  min_cells = 0,
  min_features = 0,
  verbose = TRUE
)
```

Arguments

model	GEDI model object (trained)
M	Original count matrix (optional). If not provided, will use back-transformed imputed values as approximate counts.
project	Character, project name for Seurat object (default: "GEDI")
assay	Character, name for the main assay (default: "RNA")
use_imputed	Logical, whether to add imputed data as separate assay (default: TRUE)
add_projections	Logical, whether to add ZDB and DB projections as separate assays (default: TRUE)
add_embeddings	Logical, whether to add UMAP and PCA embeddings if available (default: TRUE)
min_cells	Integer, filter genes with counts in < min_cells (default: 0)
min_features	Integer, filter cells with < min_features genes (default: 0)
verbose	Logical, whether to print progress messages (default: TRUE)

Value

Seurat object with:

- RNA assay: Original or back-transformed counts

- imputed assay: GEDI imputed expression (if `use_imputed = TRUE`)
- ZDB/DB/ADB assays: GEDI projections (if `add_projections = TRUE`)
 - ZDB: Batch-corrected gene expression (genes x cells)
 - DB: Cell embeddings in latent factor space (K x cells)
 - ADB: Pathway activities (pathways x cells) - only if C matrix provided
- umap/pca reductions: Embeddings (if `add_embeddings = TRUE` and cached)
- meta.data: Sample labels and colData from GEDI model

Examples

```
# Load example data
pbmc <- SeuratObject::pbmc_small

# Train GEDI model
gedi_model <- seurat_to_gedi(pbmc, K = 15)
gedi_model$train(iterations = 10)

# Convert back to Seurat
seurat_obj <- gedi_to_seurat(
  gedi_model,
  use_imputed = TRUE,
  add_projections = TRUE,
  add_embeddings = TRUE
)

# Now can use Seurat functions
library(Seurat)
DefaultAssay(seurat_obj) <- "imputed"
seurat_obj <- FindVariableFeatures(seurat_obj)
```

`install_optional_dependencies`

List Optional GEDI Dependencies

Description

Reports which optional packages are needed and provides install commands. Does **not** install anything automatically.

Usage

```
install_optional_dependencies(which = "all", verbose = TRUE)
```

Arguments

`which` Character vector specifying which dependency groups to query. Options: "h5" (hdf5r), "umap" (uwot), "validation" (digest), or "all" (default).

`verbose` Logical, whether to print messages (default: TRUE)

Value

A named logical vector indicating which packages are installed (invisibly).

Examples

```
# Show which optional packages are missing
install_optional_dependencies()
```

list_h5_structure *List structure of H5 or H5AD file*

Description

Helper function to explore H5/H5AD file structure.

Usage

```
list_h5_structure(file_path, recursive = TRUE)
```

Arguments

`file_path` Character. Path to the H5 or H5AD file.

`recursive` Logical. List all nested groups. Default TRUE.

Value

data.frame with file structure information

Examples

```
# Round-trip: write a tiny H5AD via write_h5ad() then list its structure.
if (requireNamespace("hdf5r", quietly = TRUE) &&
    requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)
```

```

tmp <- tempfile(fileext = ".h5ad")
write_h5ad(model, tmp)
list_h5_structure(tmp)
unlink(tmp)
}

```

plot_convergence *Plot Training Convergence*

Description

Visualizes convergence of model parameters during training. Supports multiple layout styles for different use cases.

Usage

```

plot_convergence(
  model,
  layout = c("faceted", "separate", "compact"),
  params = NULL,
  log_scale = TRUE,
  smooth = FALSE,
  title = "Training Convergence"
)

```

Arguments

model	GEDI model object
layout	Character, layout style: <ul style="list-style-type: none"> "faceted": Single plot with facet_wrap (default, best for reports) "separate": List of individual plots (for interactive exploration) "compact": Two-panel plot (global vs sample-specific)
params	Character vector, which parameters to include. Options: "Z", "A", "o", "Bi", "Qi", "oi", "si", "Rk", "Ro", "sigma2". NULL means all available.
log_scale	Logical, use log10 scale for y-axis
smooth	Logical, add smooth trend line
title	Character, plot title

Value

ggplot2 object (for "faceted" or "compact") or list of ggplot2 objects (for "separate")

Examples

```
if (requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIOObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 10, track_interval = 2)
  plot_convergence(model)
  plots <- plot_convergence(model, layout = "separate")
}
```

plot_dispersion	<i>Plot Dispersion Analysis</i>
-----------------	---------------------------------

Description

Visualizes the relationship between expected and observed variance for count data. Useful for assessing model fit quality.

Usage

```
plot_dispersion(
  dispersion_df,
  show_identity = TRUE,
  point_size = 0.1,
  alpha = 0.5,
  title = "Dispersion Analysis"
)
```

Arguments

dispersion_df	Data frame from compute_dispersion() with columns: Expected_Var, Observed_Var, Sample, n, bin
show_identity	Logical, whether to show y=x identity line
point_size	Numeric, size of points
alpha	Numeric, transparency of points
title	Character, plot title

Value

ggplot2 object

Examples

```
if (requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  M <- pbmc_small@assays$RNA@counts
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = M,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)
  disp <- model$imputed$dispersion(M)
  plot_dispersion(disp)
}
```

plot_embedding

Plot Embedding with Improved API

Description

Simplified interface for plotting embeddings with automatic caching. Model is the first argument, and color_by handles metadata/genes automatically.

Usage

```
plot_embedding(
  model,
  embedding = NULL,
  color_by = NULL,
  color = NULL,
  projection = "zdb",
  color_limits = NULL,
  palette = c("blue", "lightgrey", "red"),
  randomize = TRUE,
  point_size = 0.3,
  alpha = 0.9,
  raster = FALSE,
  xlab = "Dim 1",
  ylab = "Dim 2",
  title = NULL,
  legend_title = NULL,
  verbose = TRUE
)
```

Arguments

model	GEDI model object (or embedding matrix for backwards compatibility)
embedding	Character ("umap", "pca") or Nx2 matrix
color_by	Character: "sample", metadata column, gene name, or NULL
color	Vector for manual coloring (overrides color_by)
projection	Character: "zdb" or "db" for gene expression projection
color_limits	Numeric vector c(low, high) or NULL for auto
palette	Character vector of colors for continuous scale
randomize	Logical, randomize point order
point_size	Numeric, size of points
alpha	Numeric, transparency (0-1)
raster	Logical, use rasterization for large datasets
xlab	Character, x-axis label
ylab	Character, y-axis label
title	Character, plot title
legend_title	Character, legend title
verbose	Logical, print computation messages

Value

ggplot2 object

Examples

```
if (requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)
  plot_embedding(model, embedding = "pca", color_by = "sample")
}
```

plot_feature_ratio *Plot Two-Feature Comparison*

Description

Compares two features by computing their difference or correlation in the projected space. Mathematically grounded for GEDI's log-space representation.

Usage

```
plot_feature_ratio(
  model,
  gene1,
  gene2,
  comparison = "difference",
  embedding = "umap",
  projection = "zdb",
  color_limits = NULL,
  randomize = TRUE,
  point_size = 0.3,
  alpha = 0.9,
  title = NULL
)
```

Arguments

model	GEDI model object
gene1	Character or integer, first gene name or index
gene2	Character or integer, second gene name or index
comparison	Character, type of comparison ("difference" or "correlation")
embedding	Character specifying embedding type ("umap", "pca") or a custom N x 2 matrix
projection	Character, type of projection ("zdb" or "db")
color_limits	Numeric vector c(low, high) or NULL for auto-compute
randomize	Logical, whether to randomize point order
point_size	Numeric, size of points
alpha	Numeric, transparency of points
title	Character, plot title

Details

For comparison = "difference": Computes $(Z[\text{gene1},] - Z[\text{gene2},]) * D * B$, equivalent to $ZDB[\text{gene1},] - ZDB[\text{gene2},]$. In log-space, this represents $\log(\text{gene1}/\text{gene2})$ in the original count space. Positive values indicate $\text{gene1} > \text{gene2}$, negative indicates $\text{gene2} > \text{gene1}$.

Value

ggplot2 object

Examples

```
if (requireNamespace("SeuratObject", quietly = TRUE)) {  
  pbmc_small <- SeuratObject::pbmc_small  
  model <- CreateGEDIObject(  
    Samples = pbmc_small@meta.data$orig.ident,  
    M       = pbmc_small@assays$RNA@counts,  
    K       = 3,  
    verbose = 0  
  )  
  model$train(iterations = 5)  
  gene1 <- rownames(pbmc_small)[1]  
  gene2 <- rownames(pbmc_small)[2]  
  plot_feature_ratio(model, gene1, gene2, comparison = "difference",  
                    embedding = "pca")  
}
```

plot_features

Plot Multiple Features on Embedding

Description

Efficiently plots multiple gene features on a 2D embedding using faceting. Computes projections on-the-fly without storing full ZDB matrix.

Usage

```
plot_features(  
  model,  
  features,  
  embedding = "umap",  
  projection = "zdb",  
  color_limits = "global",  
  ncol = NULL,  
  randomize = TRUE,  
  point_size = 0.2,  
  alpha = 0.9,  
  title = NULL  
)
```

Arguments

model	GEDI model object
features	Character vector of gene names or integer indices
embedding	Character specifying embedding type ("umap", "pca") or a custom N x 2 matrix
projection	Character, type of projection to compute ("zdb" or "db")
color_limits	Character ("global" for shared scale, "individual" for per-facet scale) or numeric vector c(low, high)
ncol	Integer, number of columns in facet layout
randomize	Logical, whether to randomize point order
point_size	Numeric, size of points
alpha	Numeric, transparency of points
title	Character, plot title

Value

ggplot2 object with faceted features

Examples

```
if (requireNamespace("SeuratObject", quietly = TRUE) &&
    requireNamespace("uwot", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)
  plot_features(model, c(1, 2), embedding = "pca")
}
```

plot_vector_field *Plot Vector Field from Dynamics Analysis*

Description

Visualizes vector fields showing cell state transitions. Uses binned aggregation for cleaner visualization without overplotting.

Usage

```
plot_vector_field(
  dynamics_svd,
  color = NULL,
  alpha = 1,
  n_bins = 50,
  min_per_bin = 10,
  randomize = TRUE,
  arrow_size = 0.5,
  arrow_length = 0.15,
  arrow_color = "black",
  dims = c(1, 2),
  xlab = NULL,
  ylab = NULL,
  title = NULL
)
```

Arguments

dynamics_svd	Result from model\$dynamics\$vector_field() or similar
color	Vector of length N for coloring arrows, or NULL
alpha	Vector of length N or scalar for arrow transparency
n_bins	Integer, number of bins per dimension for aggregation
min_per_bin	Integer, minimum observations required per bin
randomize	Logical, whether to randomize data order
arrow_size	Numeric, size of arrow lines
arrow_length	Numeric, length of arrow heads (in cm)
arrow_color	Character, color for arrows (if color is NULL)
dims	Integer vector of length 2, which dimensions to plot
xlab	Character, x-axis label
ylab	Character, y-axis label
title	Character, plot title

Value

ggplot2 object

Examples

```
# Build a tiny multi-sample fixture with a sample-level prior H so that
# model$dynamics$vector_field() is available.
set.seed(1)
n_genes <- 80; n_cells <- 60; n_samples <- 3
M <- Matrix::Matrix(
  matrix(stats::rpois(n_genes * n_cells, 5), n_genes, n_cells),
```

```

    sparse = TRUE
  )
  rownames(M) <- paste0("G", seq_len(n_genes))
  colnames(M) <- paste0("C", seq_len(n_cells))
  samples <- factor(rep(paste0("S", seq_len(n_samples)),
                       each = n_cells / n_samples))
  H <- matrix(c(1, 0, 0, 0, 1, 0), nrow = 2, byrow = TRUE)
  colnames(H) <- paste0("S", seq_len(n_samples))
  rownames(H) <- c("cond_a", "cond_b")

  model <- CreateGEDIOObject(Samples = samples, M = M, K = 3, H = H,
                            verbose = 0)
  model$train(iterations = 5)
  vf <- model$dynamics$vector_field(start.cond = c(1, 0),
                                   end.cond   = c(0, 1))
  plot_vector_field(vf)

```

read_h5

Read 10X Genomics H5 file

Description

Reads a 10X Genomics HDF5 file (CellRanger v2/v3 format) and converts it to a sparse dgCMatrix suitable for use with gedi R6 object.

Usage

```

read_h5(
  file_path,
  feature_format = "gene_name",
  unique.features = TRUE,
  verbose = FALSE
)

```

Arguments

file_path	Character. Path to the 10X H5 file.
feature_format	Character. Which feature identifier to use for gene names. Options: "gene_name" (default, uses feature names) or "gene_ids" (uses feature IDs).
unique.features	Logical. Make feature names unique. Default TRUE.
verbose	Logical. Print progress messages. Default FALSE.

Value

Sparse matrix (dgCMatrix) with genes as rows and cells as columns.

Examples

```

# read_h5() expects a 10x Genomics filtered_feature_bc_matrix.h5 file.
# The example body runs only when such a file is on disk and hdf5r is
# installed; otherwise it is silently skipped.
h5_file <- "filtered_feature_bc_matrix.h5"
if (file.exists(h5_file) &&
    requireNamespace("hdf5r", quietly = TRUE)) {
  expr_matrix <- read_h5(h5_file)
  expr_matrix_ids <- read_h5(h5_file, feature_format = "gene_ids")
}

```

read_h5ad

*Read H5AD file and convert to sparse matrix***Description**

Reads an H5AD file (AnnData format) and extracts the expression matrix as a sparse dgCMatrx suitable for use with gedi R6 object.

Usage

```

read_h5ad(
  file_path,
  layer = NULL,
  use_raw = FALSE,
  transpose = TRUE,
  return_metadata = FALSE,
  feature_format = "gene_name",
  verbose = FALSE
)

```

Arguments

file_path	Character. Path to the H5AD file.
layer	Character. The layer to extract from the H5AD file. Default is NULL, which reads from X (the main expression matrix). Common alternatives include "counts", "data", "scaled", etc.
use_raw	Logical. If TRUE, reads from the raw.X slot instead of X. Default is FALSE.
transpose	Logical. If TRUE, transposes the matrix so genes are rows and cells are columns (gedi format). Default is TRUE.
return_metadata	Logical. If TRUE, returns a list with the expression matrix, cell metadata (obs), and gene metadata (var). If FALSE, returns only the expression matrix. Default is FALSE.

`feature_format` Character. Which feature identifier to use for gene names when `return_metadata = FALSE`. Options: "gene_name" (default, uses `var` rownames) or "gene_ids" (uses `var$gene_ids` column). Default is "gene_name".

`verbose` Logical. If TRUE, prints detailed progress messages. Default is FALSE.

Details

This function reads H5AD files, which are the standard format for AnnData objects in Python. Compatible with `gedi` R6 class for seamless integration.

Value

If `return_metadata = FALSE`, returns a sparse matrix (`dgCMatrix`) with genes as rows and cells as columns. If `return_metadata = TRUE`, returns a list with components:

- `X`: sparse expression matrix (genes x cells)
- `obs`: data.frame of cell metadata
- `var`: data.frame of gene metadata

Examples

```
# Round-trip: write a tiny H5AD via write_h5ad(), then read it back.
if (requireNamespace("hdf5r", quietly = TRUE) &&
    requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)

  tmp <- tempfile(fileext = ".h5ad")
  write_h5ad(model, tmp)
  expr_matrix <- read_h5ad(tmp)
  unlink(tmp)
}
```

seurat_to_gedi

Convert Seurat Object to GEDI Model

Description

Extracts count data from a Seurat object and creates a GEDI model. Automatically validates that the data contains raw counts (not normalized). Handles both Seurat v4 and v5, including split layers in v5.

Usage

```

seurat_to_gedi(
  seurat_object,
  assay = "RNA",
  slot = "counts",
  sample_column = "orig.ident",
  subset_samples = NULL,
  K = 10,
  mode = "B12",
  C = NULL,
  H = NULL,
  validate_counts = TRUE,
  use_variable_features = TRUE,
  verbose = TRUE,
  ...
)

```

Arguments

seurat_object	Seurat object
assay	Character, which assay to use (default: "RNA")
slot	Character, which slot/layer to extract (default: "counts"). For Seurat v5 with split layers (e.g., counts.CTRL, counts.STIM), this will automatically detect and combine all matching layers.
sample_column	Character, column name in meta.data for sample labels (default: "orig.ident")
subset_samples	Character vector, subset to specific samples (default: NULL = all)
K	Integer, number of latent factors (default: 10)
mode	Character, normalization mode: "B12" or "Bsphere" (default: "B12")
C	Gene-level prior matrix (genes x pathways) (default: NULL)
H	Sample-level covariate matrix (covariates x samples) (default: NULL)
validate_counts	Logical, whether to validate data appears to be counts (default: TRUE)
use_variable_features	Logical, whether to subset to highly variable features (default: TRUE). If TRUE, uses genes from VariableFeatures(seurat_object).
verbose	Logical, whether to print progress messages (default: TRUE)
...	Additional arguments passed to CreateGEDIOObject()

Value

GEDIR6 object

Examples

```
library(Seurat)

# Load example data
pbmc <- SeuratObject::pbmc_small

# Basic usage
gedi_model <- seurat_to_gedi(
  seurat_object = pbmc,
  sample_column = "orig.ident",
  K = 15
)

# Train the model
gedi_model$train(iterations = 10)
```

write_h5ad

Write GEDI model to H5AD file

Description

Exports a trained GEDI model to H5AD (AnnData) format for interoperability with Python tools like scanpy. The file contains expression data, embeddings, metadata, and GEDI-specific parameters.

Usage

```
write_h5ad(
  model,
  file_path,
  X_slot = c("imputed", "projection", "original"),
  M = NULL,
  include_embeddings = TRUE,
  include_raw = FALSE,
  compression = 6,
  verbose = TRUE
)
```

Arguments

model	GEDi R6 object (must be trained)
file_path	Character. Path where the H5AD file should be written.
X_slot	Character. Which expression data to save in the main X slot: <ul style="list-style-type: none">• "imputed": Imputed expression (default, requires M matrix)• "projection": ZDB projection (always available)

	<ul style="list-style-type: none"> • "original": Original M matrix (requires M parameter)
M	Optional. Original count matrix to save when X_slot="original" or include_raw=TRUE. Must match the dimensions used during model training.
include_embeddings	Logical. Include PCA/UMAP in obsm if cached. Default TRUE.
include_raw	Logical. If TRUE, saves original M in raw.X (requires M parameter). Default FALSE.
compression	Integer. Gzip compression level (0-9). Default 6.
verbose	Logical. Print progress messages. Default TRUE.

Details

The H5AD file structure contains:

- X: Main expression matrix (based on X_slot parameter)
- obs: Cell metadata (sample IDs, colData)
- var: Gene metadata (gene IDs)
- obsm: Cell embeddings (X_gedi, X_pca, X_umap)
- varm: Gene loadings (gedi_Z, gedi_Q_mean)
- uns: Model parameters and metadata
- raw.X: Original counts (if include_raw=TRUE)

The function handles the technical details of HDF5/AnnData compatibility:

- Writes sparse matrices in CSR format
- Transposes dense matrices for Python's row-major layout
- Creates scalar string attributes (not arrays) for AnnData compatibility
- Validates M matrix identity using fingerprints

Value

Invisibly returns the file path

Examples

```
if (requireNamespace("hdf5r", quietly = TRUE) &&
    requireNamespace("SeuratObject", quietly = TRUE)) {
  pbmc_small <- SeuratObject::pbmc_small
  model <- CreateGEDIObject(
    Samples = pbmc_small@meta.data$orig.ident,
    M       = pbmc_small@assays$RNA@counts,
    K       = 3,
    verbose = 0
  )
  model$train(iterations = 5)

  # Write to a temporary file (CRAN policy: never write to the user's
```

```
# working directory in examples).
tmp <- tempfile(fileext = ".h5ad")
write_h5ad(model, tmp)
write_h5ad(model, tmp, X_slot = "projection")
unlink(tmp)
}
```

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